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Ms. Ogunbiyi:

Here are electronic copies of both attachments to the Rule 132 Declaration submitted September 8, 2009 in the above application. Perhaps on the computer screen you can enlarge the poster document so that its contents are legible.

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Integration of a Diagnostic Into an Advanced Wound Care Dressing

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Introduction

We have developed a simple, inexpensive, safe, and rapid diagnostic sensor that detects the presence of a high burden of microorganisms in wounds before there are any visible clinical signs. The sensor measures proteases produced by bacterial pathogens that have no cross-react with human tissue or human proteases produced such as cathepsin G and human extracellular elastase. The dressing sensor is simple enough to see on a color grade board to collect on the surface of the dressing. Importantly, enough of the components test for ten years, giving the researcher and clinician a 10 year shelf life in a wound, and rapid enough to change out in 30 minutes to an hour.

The diagnostic dressing sensor is incorporated into a polyester fabric in an array that could be applied on a patient in order to continuously monitor for an infection. Clinical studies will measure the performance of the dressing sensor with patients that have chronic wounds with the long term objective of correlating healing outcome with the color change on the dressing sensor. The ultimate application for the research is to make a dressing sensor that correlates to infection. This product will allow clinicians to treat a wound infection with a therapy preventing delayed wound healing and increasing the appropriate use of antibiotics (and/or topical antimicrobial) therapies.

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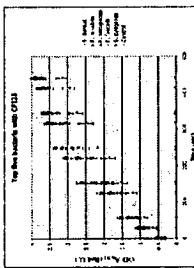
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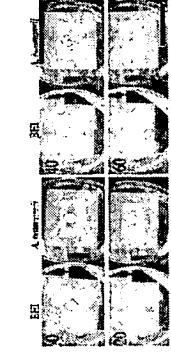
Validation of Proteases as Markers for Wound Bacteria

In vitro measurement of bacterial protease activity using biotin-polylysidine Taged CP122 microbeads. Below we demonstrate efficient processing by bacterial proteases. Cell growth medium from culture of *S. enterica* ser. Infantis, *E. coli*, *S. enterica* ser. Infantis, *P. aeruginosa* diluted to 10⁶ CFU/ml or a control sample (PBS). The media from all the bacteria (except for control) and media from a human neutrophil elastase (4 different types) were tested for control. These samples were tested on Biotin-polylysidine, CP125 beads with a 30 minute clumping time. The beads were retained by centrifugation and the clumping and increase in ELISA was determined using a microtiter plate reader and a TECAN Safire II. The sample was measured in color (450 nm) after addition of TMB substrate. We monitored the increase in color (450 nm) after addition of TMB substrate.



Testing of Dressing Sensor with Whole Blood

ECI currently has a prototype for a dressing sensor under study. The dressing sensor detects the common wound pathogens as shown in the previous figure in addition to *A. baumannii* which is an emerging pathogen in military patients that have blast induced trauma.



Dressing Sensor Time Course with *A. baumannii*. In the presence of culture media (BHT) + *A. baumannii* (diluted for 0, 20, 40, or 60 minutes). Blue color indicates the presence of the pathogen. Scale Bar = 4 cm.



Testing of ExpressoDect[®] sensors with whole blood. Uninfected white blood diluted 1/3 with PBS (A) on PBS alone (B) had no cross reactivity with the sensor. An overnight extract of *Streptococcus pneumoniae* diluted 1/3 elicits a blue color on the surface of the dressing sensor that is clearly visible in 60 minutes.

Labeling of CP125 Peptide with Blue Dye #1

We have contracted with Molecular Probes, a Division of Invitrogen (Eugene, Oregon) to synthesize 1/2 or blue dye 1 (lentiginous). The chemical structure of this food grade dye is Ammonium, ethyl[4-(1-ethyl-1-methylpropyl)amino]-alpha-(o-sulfophenyl)benzilate. 2, 5, Cyclohexadien-1,4-diene, (m-
sulfonyl)-, hydroxide, inner salt, disodium salt (CAS number 13644-45-9); as shown below. We have conjugated the indole version of blue dye #1 to peptides using a dye-to-peptide molar ratio of 1:1 in phosphate buffer pH 7.2, for 2 hours at room temperature. The labeled peptide will be removed from the unincorporated dye using a P2 column and then the dye peptide conjugate will be conjugated to HyperD CM beads using EDAC/HNHS chemistry.



Chemical structure of Ethylamine

Experimental Conclusions

1. Proteases appear to be valid markers for wound bacteria
2. The ExpressoDect[®] assay can detect bacterial proteases produced at the threshold value of 10⁶ CFU/ml *in vitro*
3. Bacterial protease activity in samples from chronic wounds correlates with to the concentration of bacteria (bacterial load) found in the wound

Integration of Assay into a Dressing Sensor

Dressing sensor prototype.

The composite dressing sensor consists of a polyethylene foam bottom layer, the CP125 - blue bead chemistry and a top membrane (Super Sorb) to collect the dye in the presence of a microbial bio-burden of 10⁶ CFU/ml.

Clinical Utility of Diagnostic Dressing Test

- * A dressing sensor would be useful in chronic wounds with a questionable infection status
 - * wounds with or without clinical signs of infection
 - * Wounds in making a correct and immediate diagnosis of local wound infection
 - * Help in treatment decision to reduce the bacterial load with a topical antimicrobial agent
 - * reduce both false negative and false positive infection diagnosis
 - * decrease the inappropriate use of antibiotics in non-infected patients
 - * allow appropriate local wound treatment to commence immediately
 - * Not be an additive step in the recommended treatment regimen
 - * substitute for a swab or biopsy where infection is confined to the wound bed
 - * topical antimicrobial agents can broad spectrum in nature
 - * test result will be useful even without the ability to determine the infecting organisms
 - * Test wounds being prepared for costly treatment or surgical options
 - * growth factor, skin grafts, or biodegradable skin

- * Test results being prepared for costly treatment or surgical options
- * growth factor, skin grafts, or biodegradable skin



ExpressoDect[®] Assay: Background

The assay detects bacteria in a wound sample by measuring the antibiotic's enzymatic (protease) activity. It measures the release of active factors (enzymes, growth factors, and growth inhibitors) from the bacteria by degrading extracellular matrix proteins, serum, derived proteins and growth factors from the bacteria. These factors are then directly measured by an enzyme assay.

**WOUND REPAIR
AND
REGENERATION**

**Rapid Measurement of Protease Activity Prevalent in
Bacteria from Wounds: A Diagnostic for Total Bioburden**

Journal:	<i>Wound Repair and Regeneration</i>
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Key Words:	proteolytic enzyme, protease, infection, bioburden, diagnostic



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3 **Rapid Measurement of Protease Activity Prevalent in Bacteria from**
4 **Wounds: A Diagnostic for Total Bioburden.**
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37 Keywords: proteolytic enzyme, protease, infection, bioburden, diagnostic
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ABSTRACT

The point-of-care diagnosis of localized wound infection is problematic because current strategies to measure bioburden rely on culture-based methods which can take at least 48 hours to obtain results. Bacterial protease activity appears to be a promising marker for the rapid and simple detection of bacteria in wound samples. Using a peptide substrate that is clipped by a broad spectrum of the bacterial species prevalent in wounds, it was shown that over 80% of wound isolates of *S. aureus*, *E. faecalis*, *S. pyogenes*, and *P. aeruginosa* produced proteases *in vitro*. Also, the major protease produced by each of these bacterial species was identified by N-terminal protein sequencing and its cleavage site within the peptide was mapped. The peptide substrate was then incorporated into a novel microbead assay format that enabled the rapid detection of proteolytic activity with a limit of detection that approached 10^5 CFU/ml for each of the 23 wound isolates measured. Clinical studies demonstrate that protease activity is present in wounds and can be measured at a level that is reflective of the appropriate bacterial bioburden (10^5 - 10^6 CFU/ml) to assist in the diagnosis of infection. Our initial clinical findings also indicate that sensitivity and specificity of detection approaches to 90% and 70% respectively.

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INTRODUCTION

Since a wound has compromised innate defenses of the skin and is open to the air, nearly all contain bacteria. However, only in certain instances are the bacteria able to overcome the immune response to slow or stall healing and cause infection. In current practice, the bacterial concentration within the wound bed is considered the most reliable marker of this event (1). The concentration of bacteria, also called bioburden, is typically assessed by quantitative microbiological analysis of a sample collected from the wound. This involves culturing a sample on rich selective media to isolate each prevalent bacterium then enumerating each species (2, 3). The clinical consensus is that a bioburden exceeding 10^5 colony-forming units per gram of wound bed tissue (CFU/g) is detrimental to wound healing and thus constitutes an infection (1, 4). This is based on seminal studies demonstrating that healing in burns and pressure ulcers, and the successful outcome of skin graft and flap procedures, depends on maintaining a bacterial level below 10^5 per gram of tissue of most bacterial species (5-12).

A significant drawback of assessing bioburden using current laboratory microbiology methodology is the time it takes to obtain results after submission of a sample (1). The culture-based methods, on which bacterial enumeration, identification, and antibiotic sensitivities depend, commonly take 48 hours or more to yield results. The identification of a molecular marker that correlates to bacterial bioburden could enable more convenient and rapid assessment of a wound and provide information at the point-of-care. Currently, no such broad-spectrum molecular marker for bioburden exists.

Virulence factors that contribute to the ability of bacteria to cause infection are obvious candidates for the broad-spectrum detection of bacteria within wounds. The measurement of enzymatic activity from virulence factors has proven to be a reliable method for the detection of bacteria in several infectious applications (13). For instance, determination of bacterial sialidase activity in vaginal samples is said to aid in the diagnosis of bacterial vaginosis. Sialidase is produced by the major causative agents of vaginosis, *Gardnerella vaginalis*,

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3 *Bacteroides spp.*, *Prevotella spp.* and various *Mobilincus spp.*, and the activity of
4 this enzyme in samples has been shown to correlate with infection as determined
5 by traditional methods (14). Another example is the detection in gingival
6 crevicular fluid of proteases produced by periodontal pathogens like
7 *Porphyromonas gingivalis* and *Tannerella forsythus* (15). This assay measures
8 bacterial proteolytic activity using a simple colorimetric peptide substrate and
9 provides information that assists in the diagnosis of periodontal disease.
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12 Proteases are produced by most of the bacterial species present in
13 wounds including *S. aureus*, *E. faecalis*, *P. aeruginosa*, and *Streptococcus spp.*,
14 and thus are a candidate marker for the wound bioburden. Secreted proteases
15 appear to be important virulence factors for several bacterial species and have
16 been shown to be produced by over 80% of *S. aureus*, *E. faecalis*, *P.*
17 *aeruginosa*, and *S. pyogenes* strains isolated from patients with a variety of
18 infection types (16-20). Also, secreted proteases from these bacteria have been
19 demonstrated to significantly contribute to virulence in animal models of infection
20 (21-25). A number of protease functions are thought to assist bacterial
21 pathogens in causing infection and would be important for bacterial persistence
22 and propagation within the wound environment. Proteases can release free
23 amino acids from host proteins to satisfy bacterial nutritional requirements,
24 process and activate bacterial surface adhesins or extracellular toxins, degrade
25 host extracellular matrix proteins, inactivate immune proteins such as cytokines
26 and immunoglobulins, and remove host cell-surface receptors (26). Another
27 potentially important role is to target and degrade antimicrobial peptides (27).
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30 This study is focused on demonstrating that bacterial protease activity is a
31 valid candidate marker for the detection of wound bacteria both *in vitro* and *in*
32 *vivo*. To demonstrate this, we developed a novel microbead-based assay
33 capable of the rapid measurement of low levels of bacterial protease activity.
34 The results indicate that protease production by bacteria isolated from wounds is
35 widespread. Also, the assay is shown to reliably detect proteases produced by
36 roughly 10^5 CFU/ml bacteria, an appropriate level to indicate infection. Our initial
37 clinical findings indicate that using the level of bacterial protease activity as a
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3 biomarker allows for detection of excessive (above the threshold of about 10^5
4 CFU/ml) bioburden with a sensitivity of 90%, and specificity of 70%.
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9 MATERIALS AND METHODS

10 Bacterial strains and growth conditions

11 Bacterial wound isolates were obtained from clinical microbiology
12 laboratories at the University of Wisconsin Hospital and Clinics (Madison, WI),
13 the Parkland Health and Hospital Systems (Dallas, TX), the University of
14 Colorado Hospital (Denver, CO), and the UCLA Medical Center (Los Angeles,
15 CA). The isolates were obtained using a variety of sampling techniques from
16 wounds of mixed etiology. Strains were initially plated on TSA plus 5% sheep
17 blood. For the measurement of protease activity, *P. aeruginosa* and *P. mirabilis*
18 were grown in nutrient broth, *S. aureus* in V8 medium (28), *E. faecalis* in brain-
19 heart infusion, and *S. pyogenes* in chopped meat medium plus glucose. Cultures
20 were grown at 37°C with shaking at 280 rpm until late logarithmic/early stationary
21 phase. Cell-free growth medium (CFGM) was prepared by centrifugation of the
22 cultures at 13,000xg for 5 minutes and filtration of the supernatant through a 0.2
23 µm filter. All clinical strains sent from the hospital microbiology labs were
24 identified by strain type only and this study was exempt from requiring IRB
25 approval.
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Proteolysis assays

Unlabeled and biotin-polyhistidine-labeled CPI2 and CPI2S (CPI2:
GAMFLEAIPMSIPC or CPI2S: GMAFLEAIPC) peptides were obtained from New
England Peptide (Gardner, MA). Protease standards endoproteinase Glu-C (V8)
and *Pseudomonas* elastase (Las B) were obtained from EMD Biosciences, La
Jolla, CA. For assays analyzing host protease activity, human neutrophil
elastase (HNE) was obtained from Athens Research and Technology, Athens,
GA. 0.5 µg/ml MMP-2 or MMP-9 (R&D Systems, Minneapolis, MN) were
activated overnight with 1 mM p-aminophenylmercuric acetate (APMA) in DMSO.

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Peptide and microbead conjugation

Unlabeled peptide substrates were conjugated to horseradish peroxidase (HRP, Roche) by first reacting HRP with sulfosuccinimidyl-4-(N-maleimidomethyl) cyclohexane-1-carboxylate (s-SMCC, Pierce) in PBS pH 7.2 for 1 hour. The resulting HRP-maleimide was purified on a P6 gel filtration column (Bio-Rad, Hercules, CA) equilibrated in PBS with 5 mM EDTA. HRP-maleimide was then coupled to a C-terminal cysteine of the peptide (CPI2 or CPI2S) by incubation for 2 hours in PBS with 5 mM EDTA, and the resulting HRP-CPI2/CPI2S was purified on a P6 column. To prepare the HRP-CPI2 or HRP-CPI2S labeled beads, a 500 μ l aliquot of Trisacryl beads (CM-Trisacryl M beads, Pall) was rinsed extensively in PBS then mixed with 40 mg of EDC (1-ethyl-3-[3-dimethylaminopropyl]carbodiimide hydrochloride) dissolved in 500 μ l 50 mM MES pH 5.2 and 2 mg of HRP-peptide in PBS buffer (roughly 500 μ l). This mixture was agitated for 3 hours at room temperature after which the bead conjugate was rinsed three times with PBS. As an alternative approach to using HRP as the reporter tag on the peptide, peptides were synthesized with a biotin and polyhistidine dual affinity tag. Synthesis of dual affinity tag-labeled CPI2s (GAMFLEAIPK(Ahx-biotin)HHHHHH) was performed by New England Peptide (Gardner, MA) utilizing Fmoc solid phase chemistry. The dual affinity tag consists of a long chain Ahx (N-e-Fmoc-e-aminocaproic acid) biotin molecule that was linked to a side chain of a lysine added to the peptide and six histidine residues (a 6xHis tag) added to the peptide C-terminus. This configuration provides sufficient spacing between the two tags to allow binding partner interactions with each to take place. The peptide was purified by reverse phase-HPLC and quality control including mass spectroscopy, HPLC, and amino acid analysis. Trisacryl CM beads (Pall Corporation, East Hills, NY) were prepared for conjugation by rinsing with Dulbecco's phosphate buffered saline (DPBS) plus 3 ppm Proclin 300 (Sigma, St. Louis, MO) three times and then with DPBS without Proclin 300 five times. After each wash, beads were collected by centrifugation. 500 mg of Trisacryl beads were then mixed with 30 mg fresh EDC (1-ethyl-3-[3-dimethylaminopropyl]carbodiimide hydrochloride) in 450 μ l of DPBS and 1 mg of

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dual affinity tag-labeled CPI2S in 50 μ l of DMSO. The entire conjugation mixture was rotated for 4 hours at room temperature. The reaction mixture was removed by centrifugation and the beads were blocked with 0.1M ethanolamine for overnight at room temperature with rotation. The conjugated beads were rinsed two times with DPBS with 0.1% polyethylene glycol 5000 (PEG 5000), one time with 0.1% Tween 20 and 0.1% PEG 5000 in DPBS for 15 minutes at room temperature, 6 more times with DPBS with 0.1% PEG 5000, another time with 0.1% Tween 20 and 0.1% PEG 5000 in DPBS for 15 minutes at room temperature, then finally rinsed 6 times with DPBS with 0.1% PEG 5000. Storage was in PBS with 3 ppm Proclin at 4°C. Bead aliquots were rinsed in 100% ethanol (2x) and placed into sterile tubes. The ethanol was removed after collection of the beads by centrifugation, and the aliquots were dried in a convection oven at 40°C and stored at 4°C.

HRP-CPI2-Trisacryl or HRP-CPI2S-Trisacryl proteolysis assay

To test correlation of the assay to protease concentration and limit of detection, the bacterial concentration (CFU/ml) in overnight cultures was estimated by A_{600} . The bacterial cells were removed, and the concentration was adjusted as indicated by dilution into fresh growth medium. Initial estimates of bacterial concentration were confirmed by dilution plating. CFGM samples were tested in triplicate 200 μ l reactions containing 100 μ l of diluted growth medium (or a PBS control) and 2 to 10 μ l HRP-CPI2-Trisacryl beads. The mixture was incubated in a 96-well filter plate for 10 minutes at room temperature then centrifuged into a catch plate to stop the reaction. 100 μ l ABTS substrate (2,2-Azino-bis-(3-ethylbenzthiazoline-6-sulfonic acid), US Biological, Swampscott, MA) was then added to each well and the increase in A_{405} was monitored for 10 minutes. The rate of color development, signifying free HRP which is a measure of the proteolytic activity, was determined for each well and the triplicate values were used to calculate average and standard deviation from the mean.

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Protease assay with dual affinity tag-labeled CPI2S-Trisacryl beads

Alternatively, dual affinity tag released by proteolysis was bound to streptavidin microtiter plates and then detected with nitrilotriacetic acid (NTA) labeled HRP as described by the manufacturer (Pierce Rockland, MD). Briefly, reactions were performed in a 150 μ l volume in a 96-well filter plate. Each reaction consisted of 100 μ l of CFGM or protease standard and 3.6 mg dual affinity labeled-CPI2S Trisacryl beads. After incubation for 30 minutes at room temperature, the 96-well filter plate was centrifuged into a 96-well plate with streptavidin-coated wells (Reactibind High Binding, Pierce). Binding was allowed for 60 minutes at room temperature with agitation and wells were rinsed three times with TBS-T (TBS buffer with 0.05% Tween 20 (polyoxyethylene sorbitane monolaureate)) for 5 minutes each at room temperature with agitation. Detection was achieved by incubation with 100 μ l of 1:1000 nitrilotriacetic acid (NTA)-labeled HRP (Qiagen) in TBS-T for 60 minutes at room temperature with agitation after which the Ni^{2+} -NTA-HRP was removed and the wells washed three times with TBS-T as described above. 100 μ l of 3,3',5,5' tetramethyl benzidine (TMB substrate SureBlue, KPL) was added and immediately transferred to a plate spectrophotometer to monitor the development of color at 650 nm for 5 minutes. The rate of color development (in mOD A_{650} /min, Vmax) was determined.

Purification of proteases able to clip CPI2

Bacteria were grown overnight in 25 ml cultures using the conditions described above and proteases were isolated from CFGM. Samples were kept at 4 °C during all purification steps. 80% ammonium sulfate $(\text{NH}_4)_2\text{SO}_4$ was added to the CFGM and the precipitated proteins were collected by centrifugation at 13,000 rpm for 20 min then solubilized in column buffer (50 mM $(\text{NH}_4)_2\text{SO}_4$ and 10 mM Tris-Cl pH 8.0). Gel filtration chromatography was performed with a Superdex 75 10/300 GL column (GE Healthcare Bioscience Corp, Piscataway NJ) equilibrated with column buffer. The protein concentration (A_{280}) of the column eluant was continuously monitored and the protease activity within each 1 ml fraction was tested by combining 20 μ l with 20 μ l of rinsed HRP-CPI2-

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3 Trisacryl beads for 5 minutes. Fractions with significant protease activity were
4 pooled, brought to a final concentration of 2.1 M (NH₄)₂SO₄, and clarified by
5 centrifugation for 7 min at 7,300 rpm. Pooled fractions (typically 7 to 8 ml) were
6 subject to hydrophobic interaction chromatography using a phenyl Hi Trap
7 column (GE Healthcare Bioscience Corp,) equilibrated with 50 mM (NH₄)₂SO₄ in
8 10mM Tris-Cl pH 8.0. Proteins bound on the column were eluted with a gradient
9 from 2.0 to 0.05 M (NH₄)₂SO₄ in 10mM Tris-Cl pH 8.0. The conductivity, protein
10 content (A₂₈₀), and CPI2 proteolytic activity of each collected fraction was
11 monitored.
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N-terminal protein sequencing

21 Purified proteases were precipitated by adding 1:10 volume of
22 trichloroacetic acid, incubating on ice for 30 min, then centrifugating for 10 min at
23 7,300 rpm. The precipitate was washed with 500 μ l ice-cold acetone then
24 suspended in 30 μ l protein sample buffer (Laemmeli Sample Buffer) and heated
25 to 100 °C for 10 min. Proteins were separated by SDS-PAGE using a 15% gel
26 and transferred to PVDF membrane (Millipore Immobilon). Proteins were
27 visualized with 0.05% Coomassie blue and excised proteins were sent for N-
28 terminal sequencing at either Commonwealth Biotechnologies Inc. (Richmond,
29 VA) or Proteos (Kalamazoo, MI). The N-terminal peptide sequences were
30 evaluated for homology to proteins in Genbank and those with the highest
31 degree of identity are reported.
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Cleavage site mapping

44 Unlabeled CPI2 (5 μ g) was incubated in a 100 μ l reaction with buffer, 1
45 μ g/ml human neutrophil elastase, 10 ng of *S. aureus* GluV8, 50 ng of *P.*
46 *aeruginosa* LasB, or cell-free growth medium from overnight cultures of *S.*
47 *aureus*, *S. pyogenes*, *E. faecalis*, *P. mirabilis*, or *P. aeruginosa* diluted to the
48 equivalent of 1x10⁷ CFU/ml. Each reaction was incubated for 60 minutes at room
49 temperature then flash frozen. A portion of each sample was then subject to high
50 sensitivity nanospray mass spectroscopy using a Thermo Finnigan LTQ ion trap
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3 mass spectrometer (Tufts University Core Facility, Boston, MA). Prominent
4 peptide fragments were identified by comparison of spectra to a buffer-treated
5 control. Cleavage sites were determined by matching the mass of an actual
6 fragment to those possible at each cleavage site.
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11 Prevalence of protease-encoding genes in wound isolates
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14 The presence of the gene encoding the CPI2-reactive protease identified
15 from purification and N-terminal sequencing was determined by gene
16 amplification using the polymerase chain reaction (PCR). Genomic DNA was
17 extracted and purified from colonies of each wound isolate using the MasterPure
18 Gram Positive DNA Purification Kit (Epicentre, Madison, WI) following
19 manufacturer's instructions. A typical 50 μ l PCR reaction consisted of 2 μ l of
20 genomic DNA, 5 μ l of 10X Thermopol buffer, 1 μ l of 10 mM dNTPs, 0.5 μ l of a
21 100 μ M stock of each primer, and 0.5 μ l Taq polymerase. The samples were
22 heated at 94 $^{\circ}$ C for 3 minutes then subject to 25 cycles of 92 $^{\circ}$ C for 30s, denaturing
23 temperature (55 $^{\circ}$ C for *P. mirabilis*, 57 $^{\circ}$ C for *E. faecalis*, 61 $^{\circ}$ C for *S. pyogenes* and
24 *P. aeruginosa*, and 65 $^{\circ}$ C for *S. aureus*) for 30 seconds, and 72 $^{\circ}$ C for 30 seconds
25 with a final extension at 72 $^{\circ}$ C for 5 minutes. Primer pairs used for PCR, listed 5'
26 to 3' for forward and reverse, were:
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29 *E. faecalis* *gelE*, GGTACAGGCATCTTGTTGG and

30 CTTGATCAACTGGATTTCCTG

31 *P. aeruginosa* *lasB*, GGAATGAACGAAGCGTTCTC and

32 GGTCCAGTAGTAGCGGTTGG

33 *S. aureus* *sspA*, GTTATATTACCAAATAACG and GCTAAATCACCTCGCCTG

34 *S. pyogenes* *speB*, GGTATCAGATTATTAAGTC and

35 CCGTTAGCGTCAAATGATCC

36 *P. mirabilis* *zapA*, CCGCAGGAAAACATATAGC and CAGAAGCCTTCATTCC

37 PCR products were analyzed by agarose gel electrophoresis and strains
38 whose DNA generated a product of the correct size were considered to contain
39 the protease gene.
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6*Clinical study design*Patient selection

The study population included patients with wounds of mixed etiology that had not healed in 60 days who provided written consent to be enrolled in the study. A university hospital wound care clinic (UMASS Medical School) served as the study setting. The study protocol was approved by the institutional review board (IRB) prior to the enrollment of subjects. Patients were screened and selected for the study based on the following criteria: at least 18 years of age, non-healing wound for 60 days, questionable infection status (i.e. not obviously infected or uninfected), not treated with an enzymatic debriding agent in the last 30 days, not receiving any investigational therapy in the last 60 days, not known to be HIV positive, and no malignancy or co-existing condition that would increase patient risk. Eligible patients were invited to enter the study, and informed consent was obtained from the subject or their legal representative.

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Sample collection and extraction

Wound tissue fluid was gathered specifically for the purpose of this study during the course of a regularly scheduled clinic appointment. Wound tissue fluid was collected from viable tissue within the wound bed using a sterile Dacron swab housed in a collection tube. If indicated in a course of treatment, non-viable tissue and cellular debris were removed by sharp debridement, surgical debridement, or cleansing of the wound with sterile saline. Tissue fluid was extracted from the wound bed using the Levine quantitative collection technique: rotating the swab over a 1 cm² area of viable tissue with sufficient pressure to express fluid (29). The swab was then returned to the collection tube and processed within 20 minutes of sampling.

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Swabs were weighed in order to estimate the volume of tissue fluid collected (difference in weight relative to the average weight of a dry swab). Tissue fluid was extracted from the swab by agitation in 1.0 ml of extraction buffer (DPBS) for 30 seconds. An aliquot of the sample (0.4 ml) was placed in a

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sterile tube and stored at 4°C prior to quantitative microbiological analysis. The
remaining 0.6 ml was flash frozen in liquid nitrogen and stored at -80°C.

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Quantitative microbiology

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A 100 µl aliquot of each swab extract was diluted 10⁻² and 10⁻⁴ in PBS and
then plated onto tryptic soy agar with 5% sheep's blood, MacConkey agar, and
chocolate agar (PML Microbiologicals Inc., Wilsonville, OR). The plates were
incubated for 48 hours at 37°C and the number of colonies of each distinct strain
was counted. The identity of each strain was determined using the Vitek II
system supplemented by standard biochemical assays when needed. These
studies were independently performed by the UMass Memorial Medical Center
Clinical Microbiology Laboratory.

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Treatment and assay of clinical samples

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300 µl of the sample was mixed with
and passed through 500 µl of a 50% slurry of sulfopropyl resin (SP; GE
Healthcare) by centrifugation in a microfuge spin filter. This treatment was found
to selectively remove the host enzyme, human neutrophil elastase (HNE) from
the tissue fluid sample (see Results Section). The bacterial protease activity in
the filtrate, referred to as SP-treated sample, was measured using dual affinity
labeled-CPI2S microbeads as described below. Samples were assessed in
groups of 10 with controls that included buffer only (negative control) and 0.1
µg/ml *S. aureus* GluV8 protease (positive control) (Worthington Biochemical).
Serial dilutions of the dual tag CPI2S peptide were also run on every assay plate
to generate a standard curve.

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Human neutrophil elastase activity in SP-treated and untreated samples
was tested using fluorogenic elastase substrate V (Methoxysuccinyl-ala-ala-pro-
val-7-amino-4-methylcoumarin, Calbiochem). Proteases in bacterial cell-free
growth medium from *S. aureus*, *E. faecalis*, *S. pyogenes*, *P. aeruginosa*, or *P.*
mirabilis did not significantly cleave this substrate (data not shown). A HNE

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3 standard curve was constructed using serial 1:2 dilutions of 100 µg/ml human
4 neutrophil elastase (Athena).
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7 The total protein concentration in tissue fluid samples was determined by
8 a Bradford assay (BioRad, Hercules, CA). A standard curve generated with
9 gamma globulin standards (0 to 300 µg/ml) was used to determine the
10 concentration (mg/ml).
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15 Protease assay with dual affinity tag-labeled CPI2S-Trisacryl beads
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17 Reactions were performed in a 150 µl volume in a 96-well filter plate.
18 Each reaction consisted of 120 µl of extracted wound fluid sample, 20 µl dual
19 affinity labeled-CPI2S Trisacryl beads from a 180 mg/ml stock, and 10 µl of
20 assay buffer (DPBS plus 0.1% PEG 5000). After incubation for 30 minutes at
21 room temperature, the 96-well filter plate was centrifuged so that the filtrate was
22 collected in a 96-well plate with streptavidin-coated wells (Reactibind High
23 Binding, Pierce). Binding was allowed for 60 minutes at room temperature with
24 agitation and wells were rinsed three times with TBS-T (TBS buffer with 0.05%
25 Tween 20 (polyoxyethylene sorbitane monolaureate)) for 5 minutes each at room
26 temperature with agitation. Detection was achieved by incubation with 100 µl of
27 1:1000 nitrilotriacetic acid (NTA)-labeled HRP (Qiagen) in TBS-T for 60 minutes
28 at room temperature with agitation after which the Ni²⁺-NTA-HRP was removed
29 and the wells washed three times with TBS-T as described above. An aliquot
30 (100 µl) of 3,3',5,5' tetramethyl benzidine (TMB) substrate (SureBlue, KPL) was
31 then added to each well and the ELISA plate was immediately transferred to a
32 plate spectrophotometer to monitor the development of color at 650 nm for 5
33 minutes. The rate of color development (in milliOD units A₆₅₀ per minute) was
34 determined from a linear portion of the data. This value was converted to the
35 mass of clipped peptide produced (ng/ml) through use of a standard curve run on
36 every plate.
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4**RESULTS****Detection of bacterial proteases *in vitro* using the CPI2 peptide**

To develop a broad-spectrum assay, a peptide substrate capable of detection of proteases produced by a number of different bacteria was needed. We utilized a peptide derived from the reactive site loop (RSL) of the mammalian protease inhibitor α 1-antitrypsin (α 1AT) since native α 1AT has been demonstrated to be cleaved by proteases from *P. aeruginosa*, *S. aureus*, *S. marcescens*, *S. pyogenes*, and *P. gingivalis* in that region (30). A 13 amino acid polypeptide derived from the α 1AT RSL was initially tested as a protease substrate. This peptide (called CPI2), corresponding to amino acids 349 to 361 of the human α 1AT RSL, was synthesized with an C-terminal lysine or cysteine GAMFLEAIPMSIPK or GAMFLEAIPMSIPC to allow labeling with a dual affinity tag (biotin and polyhistidine) or with HRP enabling measurement of cleavage. Shorter versions of this peptide (called CPI2S, GMAFLEAIP) were shown to have a reduced sensitivity to human neutrophil elastase (HNE) (see below).

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Microbead protease assay format

The CPI2 (or CPI2S) peptide was incorporated into a novel microbead-based assay format designed to improve the speed and sensitivity of bacterial protease detection (Figure 1). The main purpose of this format is to allow simple and rapid separation of clipped from unclipped peptide substrate so that the amount of clipped peptide could be measured via the tag reporter. CPI2-HRP or CPI2-Biotin-polyhistidine was attached to a chromatography microbead anchor by covalent conjugation of the available N-terminus of the peptide to carboxyl methyl groups on the Trisacryl CM bead surface. In this configuration, CPI2 serves as a hydrolyzable tether that attaches the reporter to the microbead. Bacterial protease cleavage of CPI2 should release the reporter from the surface of the microbead so that the free tag can readily be separated from the microbead-bound tag by centrifugation or filtration. The color change is measured by isolating free HRP (or indirectly through NTA-HRP) using a

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3 colorimetric substrate that correlates to the amount of peptide clipped and the
4 activity of bacterial protease in the test sample.
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7 To verify the ability of CPI2S-Biotin-polyhistidine microbeads to measure
8 bacterial protease activity, CFGM from five bacterial cultures was diluted to 1×10^8
9 CFU/ml and mixed with the bead conjugate for 30 minutes at room temperature,
10 the beads were removed by filtration, and the free tag in the filtrate was
11 measured. As seen in Figure 2A, each bacterial protease preparation released
12 the detectable dual affinity tag while media and buffer controls lacking protease
13 did not release a significant amount of reporter. This result is a clear
14 demonstration that tethering CPI2S between an affinity tag and a polymer
15 microbead did not alter the ability of bacterial proteases to clip the peptide. The
16 release of the tag was dependent on pathogen concentration (CFU/ml), as
17 shown by testing dilutions of CFGM from overnight cultures of bacterial wound
18 isolates with HRP-CPI2-microbeads. The initial concentration of the bacterial
19 culture was determined by optical density, the cells were removed, and the
20 resulting CFGM was diluted to concentrations between 1×10^8 to 1×10^6 CFU/ml in
21 fresh medium. Since the *S. pyogenes* CFGM contained much higher protease
22 activity than the other preparations, the equivalent of 1×10^7 to 1×10^5 CFU/ml was
23 tested for this species and the released HRP was diluted prior to measurement.
24 In a five-minute assay, the bacterial concentration of the sample correlated to the
25 rate of colorimetric substrate hydrolysis (Figure 2B). *S. pyogenes* CFGM had
26 roughly 10-fold more activity (taking the HRP dilution into account) than *P.*
27 *aeruginosa*, which was next most active CFGM. The remaining bacterial
28 species, *S. aureus*, *E. faecalis*, and *P. mirabilis*, produced roughly equal
29 proteolytic activity in this assay. This data clearly indicates that the response of
30 the HRP-CPI2-microbeads is dependent on the amount of protease within the
31 sample. It also shows that bacterial proteases produced by 1×10^6 CFU/ml of
32 bacteria or fewer can be effectively detected in 5 minutes.
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35 We examined the limit of detection using HRP-CPI2-microbeads with
36 several different wound isolates. Since a threshold level of 10^5 CFU bacteria per
37 gram of tissue is considered to affect healing, CFGM from wound isolates grown
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3 overnight was diluted to an *in vitro* equivalent above the threshold, 10^6 CFU/ml,
4 or below the threshold, 10^4 CFU/ml. CFGM from each wound isolate adjusted to
5 10^6 CFU/ml generated a signal that was significantly above that of a buffer
6 control in a 15-minute assay (Figure 3). At 10^4 CFU/ml only CFGM from *S.*
7 *pyogenes* and *P. mirabilis* generated a significant signal, which was consistent
8 with the high level of protease activity seen from 10^6 CFU/ml of these species.
9 This indicated that the HRP-CPI2-microbead assay detected bacteria at a level
10 above the threshold level of 10^5 CFU/ml in 15 minutes *in vitro*. The protease
11 activity detected was relatively consistent for the isolates within a particular
12 species and there was more of a difference in the average activity between
13 bacterial species. *S. pyogenes* cultures showed the most activity, followed by *P.*
14 *mirabilis*, *P. aeruginosa*, *S. aureus*, then *E. faecalis*.
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CPI2 proteolytic activity in bacteria isolated from wounds

28 The bacterial species demonstrated to produce proteases that cleave
29 CPI2 are some of the more prevalent species isolated from wound infections
30 (30). We therefore, hypothesized that bacterial protease activity toward CPI2
31 could serve as a marker for bacteria in wounds. To do so, CPI2-cleaving
32 proteases would need to have a high prevalence in wound isolates of these
33 species. This is particularly relevant in light of the demonstrated variability in
34 certain virulence traits among clinical bacterial isolates (31).
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41 To determine the pervasiveness of CPI2-cleaving proteases in bacterial
42 wound isolates, a collection of 20 strains each of *S. aureus*, *E. faecalis*, *S.*
43 *pyogenes*, and *P. aeruginosa* that were isolated from open acute or chronic
44 wounds was assembled. To ensure diversity in our strain collection, five isolates
45 of each species were gathered from four different medical centers at various
46 locations in the US. Unfortunately, one strain was incorrectly identified as *P.*
47 *aeruginosa*, resulting in its exclusion from further analysis. Protease production
48 by these clinical wound isolates was assessed and, the vast majority of the
49 wound isolates produced detectable protease activity *in vitro*. The percentage of
50 wound isolates showing CPI2 proteolysis was 90% for *S. aureus*, 80% of *E.*
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3 *E. faecalis*, 85% of *S. pyogenes*, and 94.7% of *P. aeruginosa*. This was consistent
4 with the prevalence of *in vitro* protease activity seen in isolates of these species
5 from other human infections (20, 26, 27), though somewhat higher than the
6 proteolytic activity prevalence demonstrated previously in *P. aeruginosa* isolated
7 from chronic wounds (26). As evident from the standard deviations in rate of
8 hydrolysis (Table 1), the variation of CPI2/CPI2S-dependent protease activity
9 between strains of a particular species was approximately 6-10%. Nevertheless,
10 this evidence showing that most bacterial strains from wounds produce
11 proteolytic activity toward the CPI2 substrate indicated that proteases could be
12 reliable indicators of wound bacteria. Protease activity appears to be a good
13 candidate for use as markers in a diagnostic test to detect bacteria within
14 wounds.
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Identification of bacterial proteases that clip CPI2 and characterization of their
target sites

30 To gain additional information on the proteases that may serve as
31 markers, enzymes from *S. aureus*, *S. pyogenes*, *E. faecalis*, or *P. aeruginosa*
32 that cleave CPI2 were identified. In addition, the protease from *Proteus mirabilis*
33 that clipped CPI2 was identified in order to characterize an additional gram-
34 negative organism. Proteins from the CFGM of wound isolate cultures were
35 precipitated with ammonium sulfate and the protease with significant activity
36 toward CPI2 was then purified using size exclusion and hydrophobic interaction
37 chromatography. After these purification steps, preparations were analyzed by
38 gelatin zymography and active proteases were isolated by SDS-PAGE and
39 subject to N-terminal sequencing. Each purified protein showed significant N-
40 terminal amino acid sequence homology to a known secreted proteolytic enzyme
41 from the corresponding bacterium (Table 2). *P. aeruginosa* elastase (LasB), *S.*
42 *aureus* GluV8 (SspA), and *S. pyogenes* SpeB had been previously shown to clip
43 the RSL of native α 1-AP and disrupt its inhibitory activity, so their discovery was
44 not surprising (30). However, these data provided the first evidence that *E.*
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7 *faecalis* gelatinase (GelE) and *P. mirabilis* ZapA are able to target CPI2 and
therefore the RSL of native α 1-AP.

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9 To localize the cleavage sites of the identified proteases within CPI2,
10 unlabeled CPI2 was treated with the CFGM from *S. aureus*, *P. aeruginosa*, *E.*
11 *faecalis*, *P. mirabilis*, or *S. pyogenes* and the digest was evaluated using mass
12 spectroscopy. Comparison of the sizes of the peptide fragments in digested
13 samples with untreated CPI2 enabled the identification of the cleavage products,
14 and the masses of these products allowed mapping of the target site(s) (Figure
15 4a). The CPI2 peptide cleavage map for each of the bacterial proteases is
16 shown in Figure 4b. Untreated CPI2 had a molecular mass of 1501 Da, while the
17 major digestion products were 1242 Da for *P. aeruginosa*, 853 Da for *S. aureus*,
18 1095 Da and 1242 Da for *E. faecalis*, and 853, 1101, and 1188 Da for *P.*
19 *mirabilis*. The primary target site of HNE within CPI2 was also mapped using
20 mass spectroscopy as described for the bacterial proteases. HNE produced a
21 CPI2 fragment of 1101 Da indicating that the target site was between Met/Ser
22 within CPI2, which was in a distinct location from the target sites of most of the
23 bacterial proteases tested. A schematic representation of the major peptide
24 fragments for each protease is shown in Figure 4b, similar results were obtained
25 for *S. aureus* using purified V8 protease and for *P. aeruginosa* using purified
26 LasB (not shown). Digestion of CPI2 with *S. pyogenes* CFGM produced peptide
27 fragments of 1106, 1162, and 1278 Da (not shown), which did not correspond to
28 any of the possible fragments that could result from peptide bond cleavage. One
29 possible explanation is that an enzyme in *S. pyogenes* CFGM, in addition to
30 cleavage of the peptide, is capable of modifying the mass of the CPI2 peptide
31 fragments in another manner. Common posttranslational modifications in *S.*
32 *pyogenes* include deformylation and the addition of D-alanine or thioether amino
33 acids such as lanthionine and 3-methyllanthionine. Another possibility is that *S.*
34 *pyogenes* was grown in a complex meat media which may have additional
35 peptide fragments that could mask the signal.

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37 As a next step, the prevalence of the identified CPI2-cleaving proteases
38 from each species was determined. The collection of wound isolates was
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screened for the presence of the species-specific gene encoding the CPI2-cleaving protease identified: *lasB*, *sspA*, *gelE*, or *speB* and *zapA*. Using polymerase chain reaction to specifically amplify a fragment of each of the protease genes from isolates, we determined that in each case the gene was present 100% of the time as shown in the 1% agarose gel photos in Figure 5. Therefore, the genes encoding the CPI2-targeting proteases did not appear to be a variable virulence trait in wound isolates of these species.

Time Course with V8 Protease

In order to appreciate the time dependence of the proteolytic activity for the CPI2S peptide dual affinity tag labeled beads, we performed a time course with 50 ng/ml of V8 protease standard which is comparable to the activity of *S. aureus* at 10^5 CFU per ml. Our findings indicate that within 10-15 minutes there is significant signal above the background of the uninfected media control (Figure 6). For our clinical studies reported below we conservatively used the time point of 30 minutes to measure the protease activity of actual wound fluids.

The ability of host proteases to process CPI2

A number of research studies have demonstrated that the concentrations of neutrophil proteases such as human neutrophil elastase (HNE) and gelatinase-type matrix metalloproteinase-2 and -9 (MMP-2 and MMP-9) are elevated in chronic wounds compared to acute wounds (37-38). Since HNE is the primary target of α 1-AP *in vivo* and both HNE and MMP-2 and -9 are known to cleave the reactive site loop of native α 1-AP (32-34), we investigated the ability of these proteases to clip the CPI2 peptide. Commercial preparations of HNE or activated MMP-2 or MMP-9 were diluted to physiologically elevated levels and analyzed in the protease assays. The increase in Vmax signal seen with HNE demonstrated that this protease was able to efficiently clip CPI2 comparable to the V8 bacterial protease control (Figure 7). In contrast, the signal for HNE with the shorter version CPI2S produced a signal that was much reduced without affecting the activity of V8 protease. No significant activity was

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3 seen with MMP-2 or MMP-9 with either peptide suggesting that none of the
4 residues in CPI2 are sensitive to MMPs (not shown).
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7 In Table 1, a direct comparison of the amount of HRP released from
8 CPI2S and CPI2 HRP-beads by bacterial proteases present in CFGM shows that
9 the bacterial proteolytic activities are retained while dramatically reducing the
10 sensitivity to HNE. In particular, proteases from *P. aeruginosa* or *S. pyogenes*
11 released similar amounts of HRP from CPI2 and CPI2S HRP-microbeads. *S.*
12 *aureus* and *E. faecalis* proteases were more active toward HRP-CPI2-
13 microbeads than HRP-CPI2S-microbeads, but the ratio of activity from bacterial
14 proteases and HNE showed that CPI2S appears much more selective for
15 bacterial proteases than CPI2. The bacterial protease-to-HNE activity ratios
16 reflect that CPI2S is much less sensitive to HNE relative to each bacterial
17 protease. Thus the reduced ability of HNE to clip CPI2S was not accompanied
18 by a corresponding reduction in bacterial protease activity. This truncation of the
19 peptide substrate therefore improved the specificity of detection of bacterial
20 proteases.
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Clinical study results

35 In the clinical study at UMASS Medical School, the 100 chronic wounds
36 had mixed etiologies: pressure ulcers, diabetic foot ulcers, venous ulcers, and a
37 small proportion of post-surgical and post trauma non-healing wounds (Table 3).
38 Sharp debridement was performed on wounds from 17 patients, while 12 wounds
39 were debrided surgically prior to wound fluid collection. The remaining wounds
40 were cleansed prior to sampling. Fluid collection was performed with a Dacron
41 swab using the Levine technique (29). After collection, wound fluid was extracted
42 from the swab into buffer (DPBS) and a portion of the extract was used for
43 enumeration and identification of the aerobic bacteria using standard culture
44 techniques. There were 238 bacterial strains isolated from 89 patient samples,
45 11 samples yielded no bacteria. The clinical samples had a preponderance of
46 *Staphylococcus aureus* and there was an average of 2.4 +/- 1.5 bacterial species
47 per wound falling into the range typical of other microbiological analyses (35).
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3 The 15 bacteria most frequently isolated from the clinical samples are indicated
4 in Table 4.
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7 The standard curve for HNE was performed with a fluorogenic elastase
8 substrate V (Methoxysuccinyl-ala-ala-pro-val-7-amino-4-methylcoumarin) in
9 triplicate. The range of the elastase concentration in the swab samples collected
10 using the Levine procedure was found to be 0-25.4 μ g/ml, with the mean HNE
11 concentration in 100 samples being $0.56 \pm 0.26 \mu$ g /ml and standard deviation
12 (SD) being 2.63μ g /ml. Although only a few wound fluid samples were found to
13 possess elastase at concentrations high enough to match the bacterial proteases
14 activity in terms of interaction with a CPI2 substrate (see Figure 8), and although
15 using CPI2s substantially suppresses HNE-substrate interaction, we chose to
16 find additional methods to remove HNE from the diagnostic test so that it would
17 not interfere with the assay. HNE has a very basic isoelectric point of 8.77 and
18 we determined that a cationic exchange chromatography resin such as
19 sulfopropyl (SP) would remove >80% of the HNE. The range for HNE
20 concentration in SP-treated samples became 0 to 1.6 μ g /ml, with the mean
21 value being $0.07 \pm 0.02 \mu$ g /ml and SD= 0.22μ g /ml.
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24 In addition to testing HNE activities, we measured the protease activity of
25 each clinical sample. Each reaction consisted of 120 μ l of extracted wound fluid
26 sample, 20 μ l of dual affinity labeled-CPI2S Trisacryl beads from a 180 mg/ml
27 stock, and 10 μ l of assay buffer (DPBS plus 0.1% PEG 5000). After
28 centrifugation, the detection of released dual affinity tag was performed in a
29 streptavidin ELISA plate as described in the methods section. The data collected
30 was a measurement of protease activity in each of the samples (mOD/min) which
31 was then converted to total nanograms of peptide cleaved using a standard
32 curve of CPI2S dilutions. A subsequent analysis found no apparent correlation
33 between the level of HNE and the amount of CPI2S clipped peptide in SP-treated
34 samples ($R^2=5 \times 10^{-3}$, $F=0.53$, $p=0.47$). Thus, we conclude that our detection
35 technique is specific for bacterial proteases.
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38 The microbiology results were obtained from the UMASS Medical School
39 Clinical Microbiology Lab and the total bioburden (CFU/ml) was compared to the
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protease activity/nanograms of peptide clipped for each sample. Statistical
analysis of the clinical data demonstrated a significant correlation between
proteolytic activity and total bioburden ($p < 10^{-4}$).

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10 A threshold level of bioburden to discriminate between infected and
11 uninfected wounds is believed to be in a range of 10^5 - 10^6 CFU/ml (1, 12). The
12 Wound Healing Society recommends that all chronic and acute wounds should
13 be treated at a bioburden level of $\geq 1 \times 10^6$ CFU/ml for most bacteria. In our
14 attempt to reliably distinguish between infected and uninfected wounds on the
15 grounds of exhibited proteolytic activity, we built a family of Receiver Operating
16 Characteristics (ROC) that gave a relationship between sensitivity and false
17 positive rate of detection for every particular choice of bioburden threshold
18 discriminating between infected and uninfected wounds. The greater the area
19 under the ROC curve (0.5 corresponds to a test of no diagnostic value, while 1 is
20 equivalent to a golden standard), the more robust a diagnostic test is believed to
21 be. The choice of bioburden cutoff at 3×10^5 CFU/ml yields an optimal ROC to
22 maximize an area under the curve (AUC), with the bootstrap estimate for AUC
23 being 0.84 ± 0.06 (SAS version 9.0, SAS Institute Inc., Cary, NC). Similarly, the
24 Leave-one-Out method of cross-validation yields $AUC = 0.84 \pm 0.05$ ($\chi^2 = 267.7$,
25 $p < 10^{-4}$), while the most conservative method of Split-Sample validation results in
26 $AUC = 0.84 \pm 0.09$ ($\chi^2 = 85.6$, $p < 10^{-4}$). The mean activity of uninfected samples
27 (bioburden $< 3 \times 10^5$ CFU/ml) is found to be 26.9 ± 8.7 ng of peptide clipped per
28 sample, which is much lower than the mean activity of infected samples
29 (bioburden $\geq 3 \times 10^5$ CFU/ml) found to be 77.1 ± 6.2 ng of clipped peptide. The
30 comparison of the mean proteolytic activity of infected and non-infected clinical
31 samples (Figure 9) proves their significant difference, with a Wilcoxon rank-sum
32 test (equivalent to the Mann-Whitney U-test) giving us the two-tailed asymptotic
33 p-value for the probability of the difference occurring by chance alone as $p < 10^{-4}$.
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We found that limiting the collection volume to 0.1 ml of wound fluid,
suppresses sample-to-sample variability and helps us to better distinguish
between infected and uninfected wounds in terms of protease activity.

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The choice of activity cutoff in the range from 33 ng to 50 ng of peptide clipped/0.1 ml yields detection sensitivity of 89.6% and a specificity of 69.6% relative to the reference standard of quantitative microbiology (Figure 10). When discriminating between "infected" and "uninfected" wounds regarding the level of their activity measured per 0.1 ml of sample, a Wilcoxon rank-sum test gives us the two-tailed exact p-value for the probability of rejecting the null hypothesis by mistake as $p=6.78\times10^{-8}$.

In Fig. 11 (a) we sorted all 100 wounds according to their level of bacterial protease activity (below/above 45 ng of clip per 0.1 ml) and according to their bioburden (below/above 3×10^5 CFU/ml). There is a two orders of magnitude difference in mean activity values between "active" and "inactive" groups vs. only 5 times difference in mean activity values between "infected" and "uninfected" groups.

We speculate that some "false positives" of detection (high level of protease activity measured in wounds with a low bacterial load) come from intrinsic enzymatic activity associated with a dead tissue, present in non-debrided wounds. While bacteria are mainly immobilized within the dead tissue, the enzymes they produced are mobile and able to contribute to the net protease activity found in the wound fluid. The analysis of data obtained from all 29 debrided wounds in our study is depicted in Fig. 11b.

Whereas a difference in the mean activity values between "active" and "inactive" groups stays approximately the same, the difference in the mean activity values between "infected" and "uninfected" groups increases substantially. In terms of ROC analysis, an estimate for the area under the curve gives much larger $AUC=0.92\pm0.05$. The same choice of activity threshold (within a range of 33 to 50 ng/0.1 ml) yields marginally better sensitivity of 92.9 % but much better specificity of 80%. If specificity of detection is of great concern (a low prevalence case), it can be increased even more at the penalty of lower sensitivity, namely, setting activity threshold in the range of 100 to 130 ng /0.1 ml results in 85.7% sensitivity and 86.7 % specificity of detection.

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DISCUSSION

The risk of wound infection depends on a combination of bacterial and patient factors. Patient factors include the underlying disease state, immune status, and local influences: wound location, size, depth, oxygen perfusion, and the presence of foreign material. Bacterial factors include the bacterial concentration, the number of bacterial species and the virulence potential of microbes, bacterial biofilms, and synergistic interactions (36). Despite this basic understanding, there is relatively little information regarding the virulence mechanisms that assist bacteria in causing wound infection. The data presented here indicate that proteolytic enzyme production is a common virulence trait among bacterial wound isolates. Consequently, bacterial protease activity has the potential to be a diagnostic marker for the detection of bacteria in wounds.

The first goal of this study was to identify a peptide substrate able to simultaneously detect proteases produced by the different bacterial species found in wounds. We demonstrated that CPI2, a peptide substrate that is the reactive site loop (RSL) of the wide spectrum protease inhibitor human alpha-1 antitrypsin (α 1AT), was clipped by secreted proteases from five predominant bacterial species prevalent in wounds, namely *S. aureus*, *E. faecalis*, *S. pyogenes*, *P. aeruginosa*, and *P. mirabilis*. In similar studies, CFGM from overnight cultures of *Serratia marcescens*, *Enterobacter cloacae*, *Morganella morganii*, *Acinetobacter baumanii*, and *Stenotrophomonas maltophilia* also clipped the CPI2 peptide (data not shown). This led to the conclusion that CPI2 is a capable broad-spectrum substrate for bacterial proteases. This was consistent with a previous report of a multi-step assay used to detect protease activity in unknown samples based on the broad susceptibility of α 1AT to proteolytic attack. One major improvement over that report is the use of a target peptide representative of the α 1AT RSL, rather than the whole protein, and its incorporation into a simple, rapid assay format.

Purification of the primary proteases that target CPI2 and mapping of their target sites demonstrated that CPI2 contains a number of residues surrounding each of the identified bacterial protease target sites. This includes at a minimum

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the P'2, P'1, P1, and P2 residues, which gives bacterial proteases adequate spacing to recognize the target and efficiently clip the peptide. The identified target site for *S. aureus* GluV8 in this study is identical to that determined previously for native α 1AT (37) and consistent with the P1' glutamic acid preference of GluV. In contrast, the mapped site for *P. aeruginosa* LasB cleavage, Met/Phe, is different than that previously reported for native α 1AT. Morihara et al. (1979) previously identified Pro/Met as the LasB cleavage site (38). This discrepancy may be due to the different context of the target site within the soluble CPI2 peptide and within the RSL in the α 1AT protein, which could alter the preferred site of cleavage.

Another important goal of this study was to examine the prevalence of protease production by bacterial wound isolates. The proteases identified to clip CPI2 were shown to be invariable virulence traits in 20 wound isolates of *S. aureus*, *E. faecalis*, and *S. pyogenes*, 19 wound isolates of *P. aeruginosa* and 3 wound isolates of *P. mirabilis* from four clinical locations within the US. This was demonstrated by species-specific PCR amplification indicating that each wound isolate contained the gene of interest. When expression of these genes was determined by measuring CPI2-dependent protease activity in the wound isolates, >80 to 95% produced detectable amounts of protease *in vitro*. In addition, 3 of 3 *P. mirabilis* wound isolates displayed activity (data not shown). The differences evident in protease activity among species and between species are most likely due to variation in the protease expression levels that result in a different concentration of proteases in the CFGM from each isolate. Also, each bacterial enzyme may have a different specific activity towards CPI2.

Another significant aspect of this work is the demonstration of a simple, colorimetric assay able to provide a rapid indication of protease activity. The primary reagent for this assay is the reporter-peptide substrate-microbead conjugate produced by covalent attachment of a peptide labeled with an affinity tag to the modified surface of a polymer bead (Figure 1). This configuration allows simple and rapid separation of reporters released from beads due to peptide cleavage and those that remain on the bead attached to unclipped

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4 peptide. Similar constructs have been used with peptide microarrays to study
5 protease catalytic activity and cross-reactivity though these mostly utilize a
6 fluorescent reporter. The major potential concern with this format, access of the
7 peptide tethered-bead substrate to bacterial proteases, does not appear to be a
8 significant issue since CFGM from multiple species clipped CPI2 and CPI2S and
9 released detectable HRP or the dual affinity tag (Figure 2a+b). Since this
10 microbead format could be used with virtually any peptide substrate, the assay
11 could be used to rapidly measure any protease activity with a specificity
12 determined by the peptide substrate. Also, our current approach produces a
13 colorimetric readout, replacement of the affinity tag with another reporter, such as
14 a fluorescent or colored dye would allow other types of output. Consequently,
15 the microbead-based assay could have utility in a number of laboratory or clinical
16 diagnostic applications.
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19 The specificity of the HRP-CPI2-microbead assay for bacterial proteases
20 is critical for a clinically useful test to detect bacteria in wound samples. The
21 CPI2 peptide is susceptible to cleavage by the inflammatory protease HNE but
22 not MMP9, which can be present at high levels in a subset of chronic, or non-
23 healing, wounds (39). Since elevated HNE could cause protease activity to be
24 detected in the absence of bacterial proteases, CPI2 was altered to reduce
25 cleavage by HNE relative to bacterial protease. Through peptide mapping of the
26 target sites of bacterial proteases and HNE within CPI2 (Figure 4a+b), four C-
27 terminal amino acids that contained the HNE target site were removed to
28 generate CPI2S, while the N-terminal portion of the peptide containing the
29 bacterial protease target sites was left intact. This dramatically improved the
30 bacterial specificity of the peptide substrate without significantly raising the limit
31 of detection. CPI2S showed a 93% reduction in cleavage by HNE and an
32 increased selectivity (represented by bacteria protease-to-HNE ratio) toward
33 bacterial proteases. This new peptide substrate, in addition to a sample filtration
34 through the SP-resin to selectively bind and reduce HNE content, was used to
35 detect bacterial protease activity in specimens from wounds and correlate this
36 activity to bacterial content.
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3 We found that bacterial protease activity in samples from chronic wounds
4 correlates well with the concentration of bacteria (bioburden) found in the wound.
5 An initial 100 patient clinical study suggested that our assay can have a
6 sensitivity of about 90% and a specificity of about 70% for detection of wound
7 bioburden exceeding the threshold between 10^5 and 10^6 CFU/ml. We believe
8 one source of false positives compromising specificity of detection to be an
9 enzymatic activity associated with a dead tissue. Actually, the Wound Healing
10 Society recommends wound debridement to be performed, when necrotic tissue
11 is present. For debrided wounds, our assay demonstrated better performance,
12 with a sensitivity close to 93% and a specificity of 80%. If specificity of detection
13 is of primary concern, it can be increased by adjusting the assay detection
14 threshold, at the cost of lower sensitivity, to yield approximately 86% sensitivity
15 and 87% specificity of detection.
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18 In summary, the studies presented here demonstrate the validity of using
19 bacterial protease activity as a biomarker for the detection of wound bacteria.
20 These enzymes are the first biochemical marker to serve this purpose and the
21 creation of an assay to rapidly and simply measure their activity is a critical step
22 in the development of a rapid test that will help establish an accurate point-of-
23 care infection diagnosis and treatment plan for patients with wounds. A marker
24 such as protease activity depends on both the concentration of bacteria and their
25 ability to express and secrete proteases. Thus protease activity has the ability to
26 provide an indication of both the level and virulence potential of bacteria within
27 wounds and could offer an improved measure of infection status compared to the
28 current standard of bacterial concentration alone. The ultimate goal of this
29 project is to compare the correlation of bacterial protease activity and bacterial
30 concentration to healing outcomes.
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5 **Table 1:** Comparison of protease cleavage of CPI2S and CPI2

	# of Strains with Activity	CPI2 Activity Vmax (OD _{405nm})	CPI2S Activity Vmax (OD _{405nm})	bact/HNE ratio CPI2	bact/HNE ratio CPI2S
HNE	NA	106 +/- 6.6	6.9 +/- 1.8	n/a	n/a
<i>S. aureus</i>	18/20 (90%)	188 +/- 20	82.9 +/- 5.0	1.8	12.1
<i>E. faecalis</i>	16/20 (80%)	80.7 +/- 7.5	44.9 +/- 0.9	0.8	6.5
<i>S. pyogenes</i>	17/20 (85%)	25.7 +/- 2.5	18.8 +/- 0.9	0.2	2.7
<i>P. aeruginosa</i>	18/19 (94.7%)	194 +/- 13.4	152 +/- 31.1	1.8	22.2

22 **Table 2:** Identification of bacterial proteases able to cleave CPI2

	comparison	match
<i>P. aeruginosa</i>	AEAGGPGGNQ AEAGGPGGNQ	elastase (LasB)
<i>S. aureus</i>	VILPNNDRHQ VILPNNDRHQ	glutamyl endopeptidase (GluV8, SspA)
<i>S. pyogenes</i>	GTAEIKQPVVKSL LDS GTAEIKQPVVKSL LDS	streptopain (SpeB)
<i>E. faecalis</i>	VGSEVTLKNS * VASEVTLKNS	coccolysin (GelE)
<i>P. mirabilis</i>	NYSTKVLP * NYGTVKLP	mirabilysin (ZapA)

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5**Table 3:** Chronic Wound Etiology6
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Wound type, by clinical diagnosis:10
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21 pressure ulcers
20 diabetic foot ulcers
17 venous ulcers, some patients also diabetic
29 etiology not indicated/not known
13 post-surgical or post-trauma non-healing wounds
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20**Table 4:** Composition of Bacteria Found in Chronic Wounds

	#	% of isolates	% of patients
<i>Staphylococcus aureus</i>	43	18.1	43
<i>Corynebacterium spp.</i>	41	17.2	41
<i>Staphylococcus spp. (not aureus)</i>	32	13.4	32
<i>Enterococcus species</i>	16	6.7	16
<i>Pseudomonas aeruginosa</i>	14	5.9	14
<i>Proteus mirabilis</i>	11	4.6	11
<i>Escherichia coli</i>	9	3.8	9
Beta-hemolytic Strep Group B	8	3.4	8
Beta-hemolytic Strep Group G	8	3.4	8
<i>Acinetobacter baumannii</i>	7	2.9	7
<i>Viridans streptococci</i>	6	2.5	6
<i>Klebsiella pneumoniae</i>	6	2.5	6
<i>Morganella morganii</i>	5	2.1	5
<i>Stenotrophomonas maltophilia</i>	5	2.1	5
<i>Klebsiella oxytoca</i>	4	1.7	4

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FIGURE LEGENDS

Figure 1: Diagram of the microbead format for measuring protease activity. The CPI2 peptide is covalently conjugated to a Trisacryl microbead (roughly 70 μ m in size) via the N-terminus and to a tag or dual tag via a C-terminal cysteine. The peptide acts as a tether that upon cleavage by bacterial proteases releases the detectable tag from the microbead. Free cleaved tag can then be separated from tags that are bound to microbeads by filtration or centrifugation and measured directly or using a colorimetric substrate.

Figure 2: *In vitro* measurement of bacterial protease activity using biotin-polyhistidine tagged CPI2 microbeads and HRP-CPI2-microbeads.

A) Demonstration of efficient processing by bacterial proteases. CFGM from cultures of *S. aureus*, *E. faecalis*, *S. pyogenes*, *P. mirabilis*, or *P. aeruginosa* adjusted to 10^8 cfu/ml or a control sample (PBS). The media from all the bacteria grow ups (4 different types) was tested as controls and gave the same signal as PBS. Data is not shown to simplify the graph. These samples were tested on Biotin-polyhistidine-CPI2S beads with a 30-minute cleavage time. The beads were removed by centrifugation and free dual tag was measured in an ELISA assay using streptavidin -coated plates and NTA-HRP. Signal was measured by monitoring the increase in color ($A_{650\text{ nm}}$) after addition of TMB substrate.

B) Correlation of signal tag release to bacterial protease concentration. CFGM from overnight cultures of *S. aureus* (red), *E. faecalis* (purple), *S. pyogenes* (blue), *P. aeruginosa* (green), or *P. mirabilis* (black) of known concentration was diluted to 1×10^5 to 1×10^8 CFU/ml then assayed using HRP-CPI2 microbeads for 10 minutes. *S. pyogenes* was diluted 1:10 for this study because the beads detect it at a lower level. The amount of signal tag released was determined by the rate of color development using ABTS substrate. The signal demonstrated a concentration-dependent increase with bacterial concentration in the sample.

Figure 3: *In vitro* proteolytic activity of CFGM from bacterial wound isolates diluted to 10^6 CFU/ml (black) or 10^4 CFU/ml (grey). Diluted sample or a buffer control was mixed with HRP-CPI2-Trisacryl beads for 10 minutes and the amount

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3 of HRP released was measured. The assay was performed in triplicate and bars
4 represent the limits of the standard deviation from the mean.
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9 **Figure 4:** Mass spectroscopy analysis of CPI2 fragments

10 (A) Mass spectroscopy data for limited proteolysis of the CPI2 peptide with *P.*
11 *aeruginosa*, *E. faecalis*, *P. mirabilis*, *S. aureus*, HNE and undigested control
12 peptide (MW 1501).

13 (B) Protease cleavage sites within the CPI2 peptide. Unlabeled CPI2 was
14 exposed to human neutrophil elastase (HNE) or CFGM from *S. aureus* (SspA),
15 *E. faecalis* (GelE), *S. pyogenes* (SpeB), *P. aeruginosa* (LasB), or *P. mirabilis*
16 (ZapA) then analyzed by mass spectroscopy. The locations of predominant
17 cleavage sites were determined by comparison of the mass of the proteolytic
18 fragments to possible cleavage products.

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26 **Figure 5:** PCR of protease genes found in *S. aureus* (V8), *P. aeruginosa* (LasB),
27 *E. faecalis* (GelE), *S. pyogenes* (SpeB), and *P. mirabilis* (ZapA). The presence of
28 the protease genes V8, LasB, GelE, SpeB, and ZapA was determined by
29 polymerase chain reaction (PCR) by using primers specific to each gene to
30 amplify a 290-320 base pair product for each gene. Genomic DNA was extracted
31 from cell scraping off blood agar plates for each sample. The MasterpureTM
32 Gram Positive (Epicenter Biotechnologies, Madison WI) or the PrepmanTM
33 (Applied Biosystem, Foster City, CA) DNA purification kits was used to isolate the
34 DNA, 2ul of each genomic prep was used as a template for the PCR reactions. In
35 all case the genes were found in 100% of strains examined suggesting that these
36 are essential for virulence factors in each wound pathogen. All PCR reactions
37 were performed for 25 cycles with a melting temperature of 92°C, anneal at 50°C
38 and extension at 72°C in a MyIQ Single Color Real-Time PCR detection system
39 (Bio-Rad, Hercules, CA).

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52 **Figure 6:** A time course study was performed on Biotin-polyhistidine-CPI2S
53 beads with a 5,10, 15 and 30 minute cleavage time. Purified V8 protease
54 standard (EMD Biosciences) at 50 ng/ml was used to cleave the peptide. The
55 beads were removed by centrifugation at the designated time and free dual tag
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was measured in an ELISA assay using streptavidin -coated plates and NTA-HRP. Signal was measured by monitoring the increase in color (650 nm) after addition of TMB substrate. The time dependence of cleavage is shown in the graph versus a control of PBS buffer.

Figure 7: Improvement of substrate peptide to reduce cleavage by host proteases. Biotin-polyhistidine CPI2 peptide (circles) on beads shows a concentration dependent increase in signal from human neutrophil elastase (HNE) tested in the 0-1 μ g/ml range. The peptide on beads was exposed to HNE concentrations for 15 minutes and then the beads were filtered out of solution. The released tag was measured by ELISA and TMB color development. The graph shows the Vmax at 650 nm for each concentration. The HNE cleavage site was removed from the CPI2 peptide in a newer version of the substrate: CPI2S. CPI2S shows reduced HNE activity (triangles) tested in the same manner in the same concentration range. Also shown is the reactivity of CPI2 and CPI2S peptide to purified V8 bacterial protease (diamonds and squares respectively) in the 0-1 μ g/ml range which is not greatly affected by the removal of the elastase cleavage site. MMP 2 and MMP 9 (0.5 μ g/ml) were also tested on the CPI2 peptide. The results (not shown) yielded the same signal as a control indicating no CPI2 cleavage.

Figure 8: Human Neutrophil Elastase (HNE) was efficiently removed cation exchange (SP) resin. The samples that had the most elastase activity are shown before and after treatment with 500 μ l of a 50% slurry SP cation exchange resin. We determined that SP resin is very effective at removing HNE from a wound, suggesting that this method could be used to improve the signal to noise of the diagnostic.

Figure 9: Comparison of the mean proteolytic activity of infected and non-infected clinical samples. Error bars represent the 95% confidence interval of the mean.

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3 **Figure 10:** (a) The protease activity was tested on 100 clinical samples from
4 patients with non-healing wounds (>60days) attending the UMass Medical
5 School Wound Clinic. We only considered patients with questionable infection
6 status. This data shows a scatter plot of total CFU/ml vs. ng of peptide clipped
7 normalized to 0.1 ml. Wounds that were presumed infected or truly uninfected by
8 clinical signs were excluded from the study as well as any patient that had been
9 treated with an enzymatic debriding agent for 30 days. The 45 ng of peptide
10 cutoff was chosen to distinguish between infected ($\geq 3 \times 10^5$ CFU/ml) and
11 uninfected ($< 3 \times 10^5$ CFU/ml) samples. (b) The clinical data represent: 69 true
12 positives, 16 true negatives, 8 false negatives, and 7 false positives. The
13 bacterial protease activity in samples from chronic wounds correlates with the
14 concentration of bacteria (bioburden) found in the wound. The sensitivity of the
15 test is 89.9% with a specificity of 69.6% and an overall accuracy of 85%.

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17 **Figure 11:** Comparison of the mean proteolytic activity of “active” vs. “inactive”
18 wound samples (based on the level of bacterial protease activity: above/below 45
19 ng of clip per 0.1 ml) and mean proteolytic activity of “infected” vs. “uninfected”
20 wound samples (based on the bioburden: above/below 3×10^5 CFU/ml). Activity is
21 measured in nanograms of peptide clipped. Error bars represent the 95%
22 confidence interval of the mean. (a) All 100 wound samples ; (b) Only 29
23 debrided wound samples.

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FIGURES

Figure 1

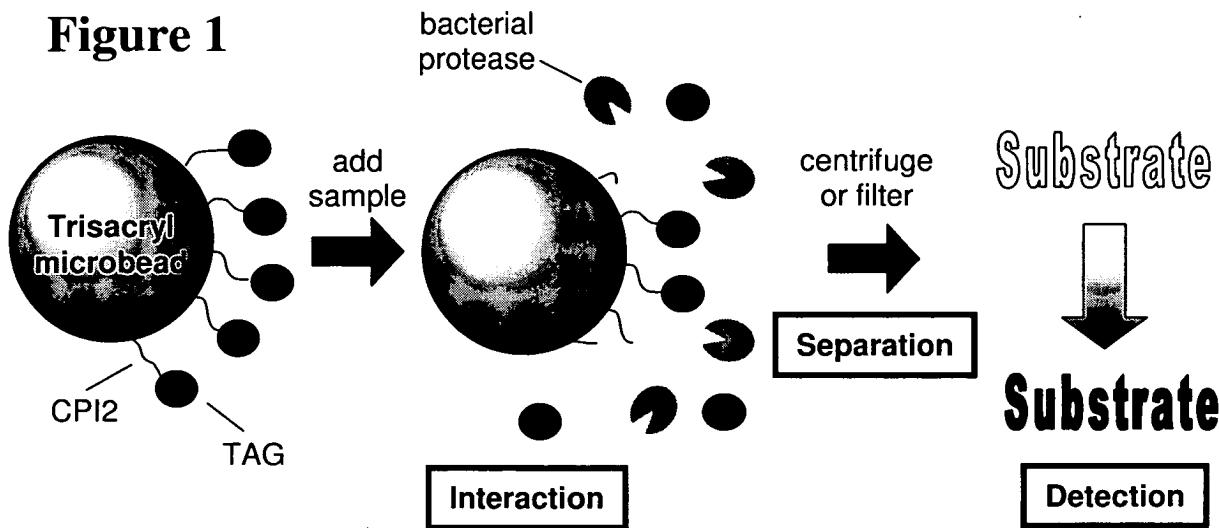


Figure 2 (a)

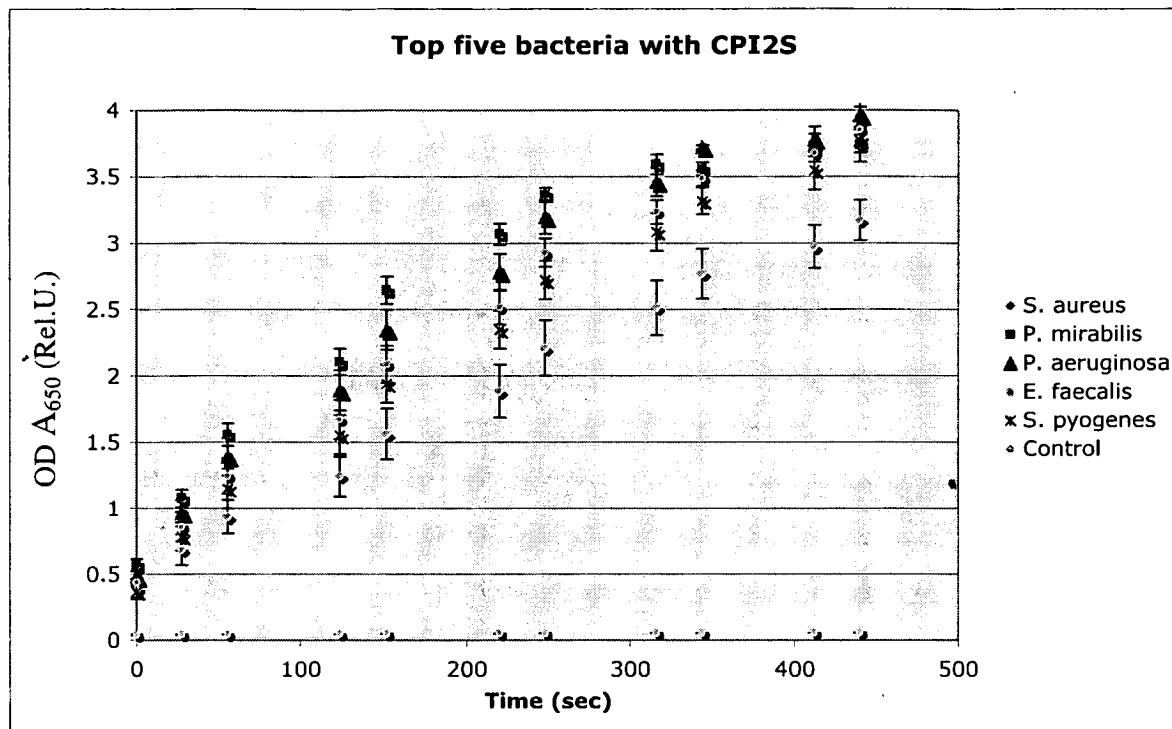


Figure 2 (b)

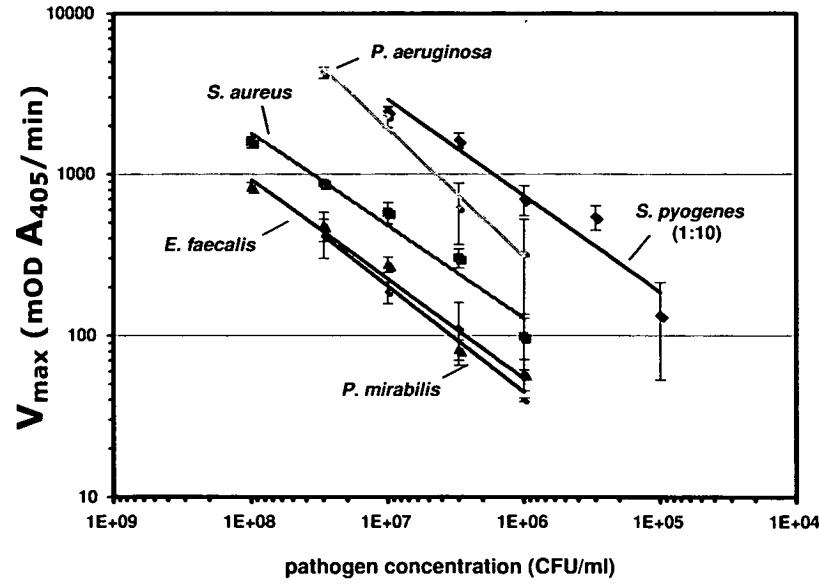


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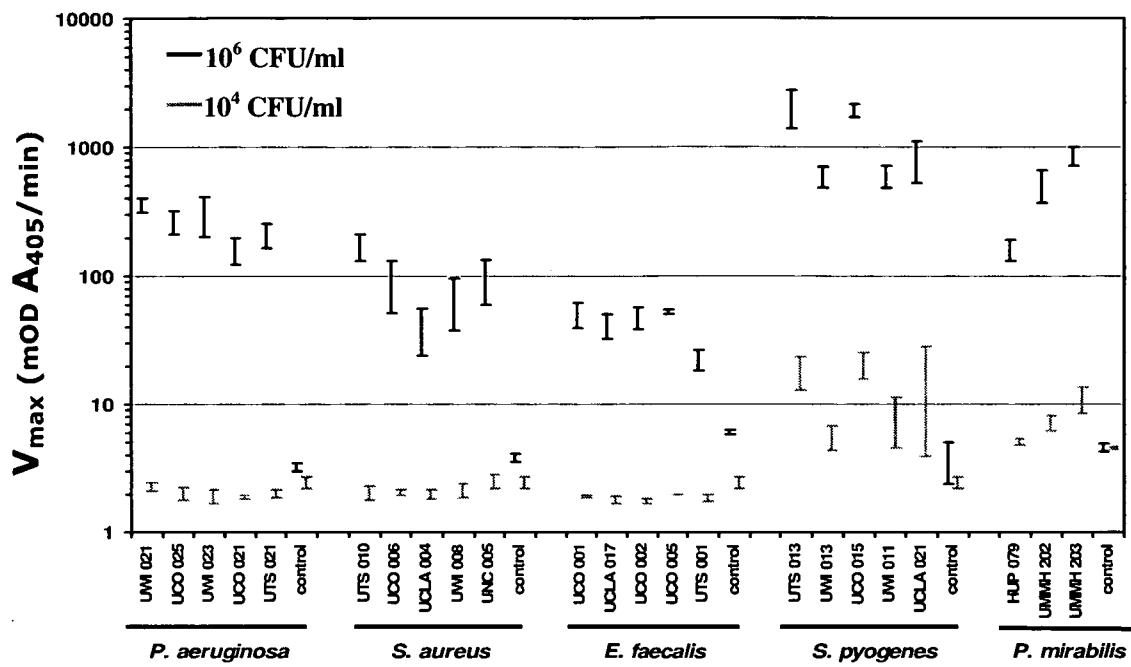
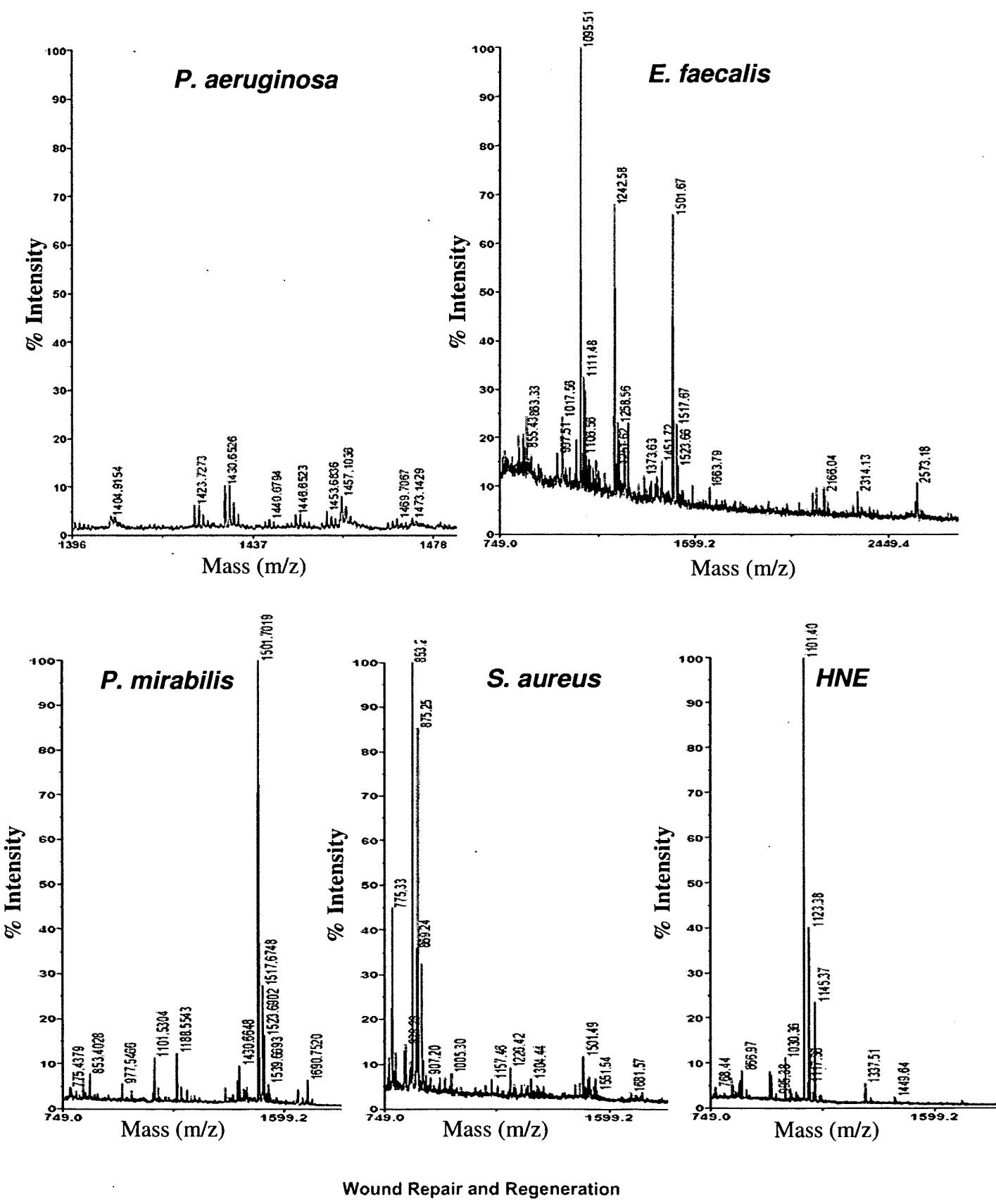


Figure 4 (a)



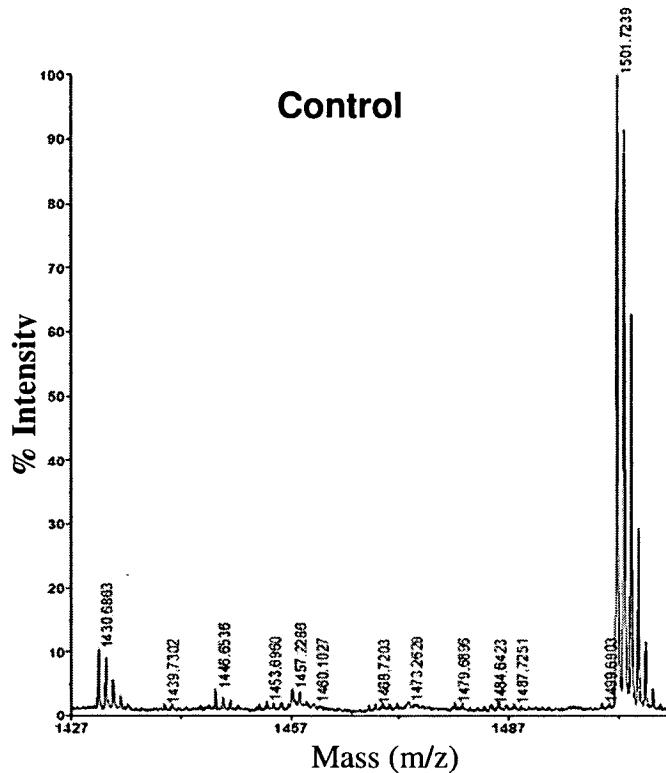
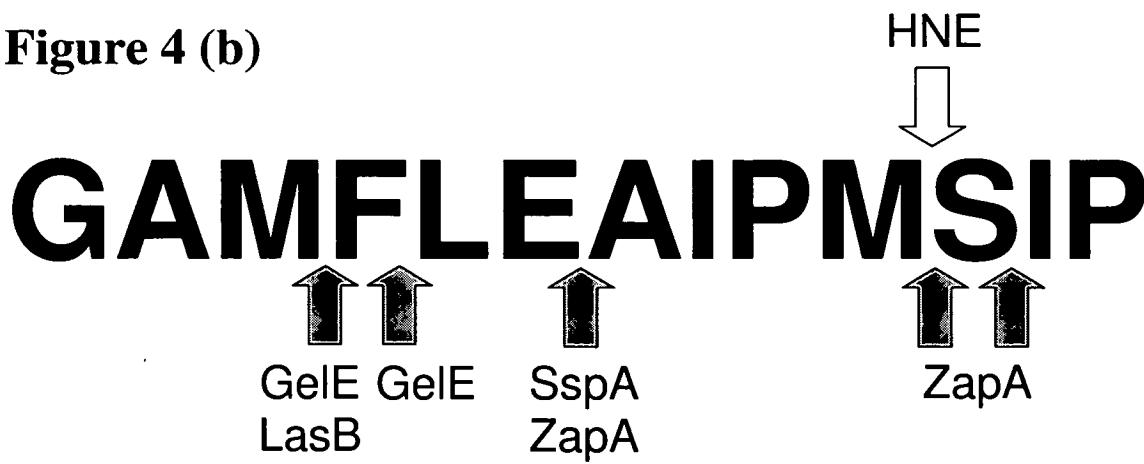


Figure 4 (b)



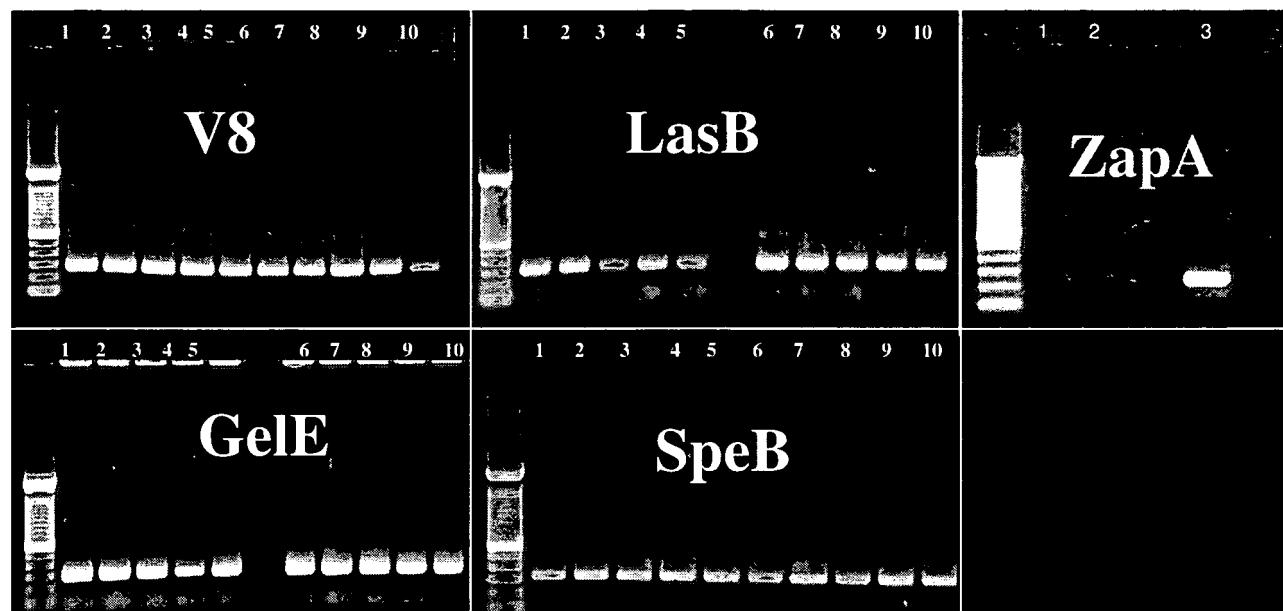
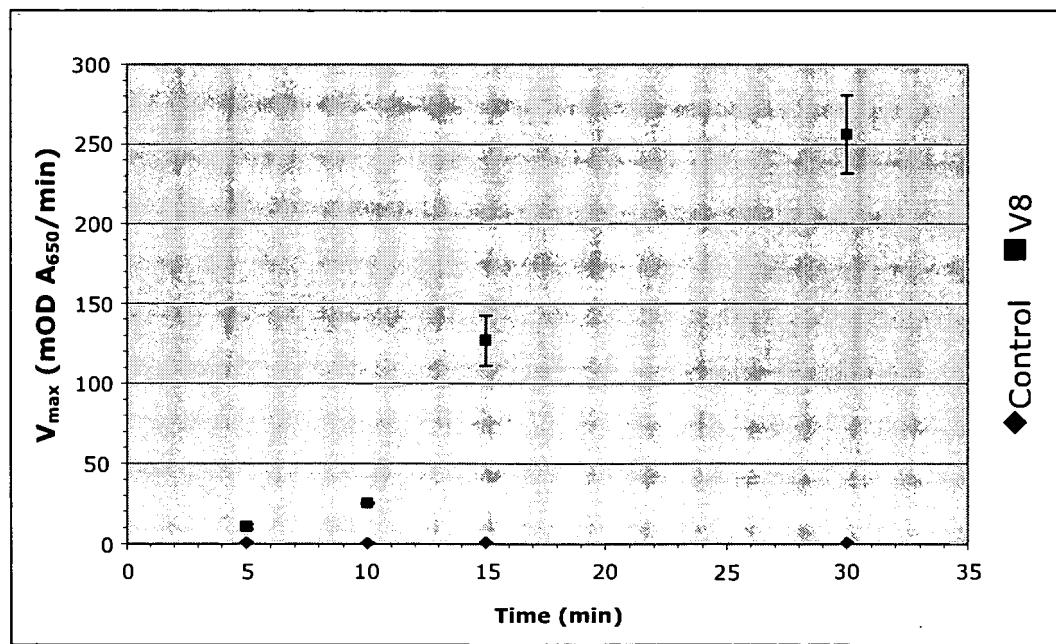
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6**Figure 5****Figure 6**

Figure 7

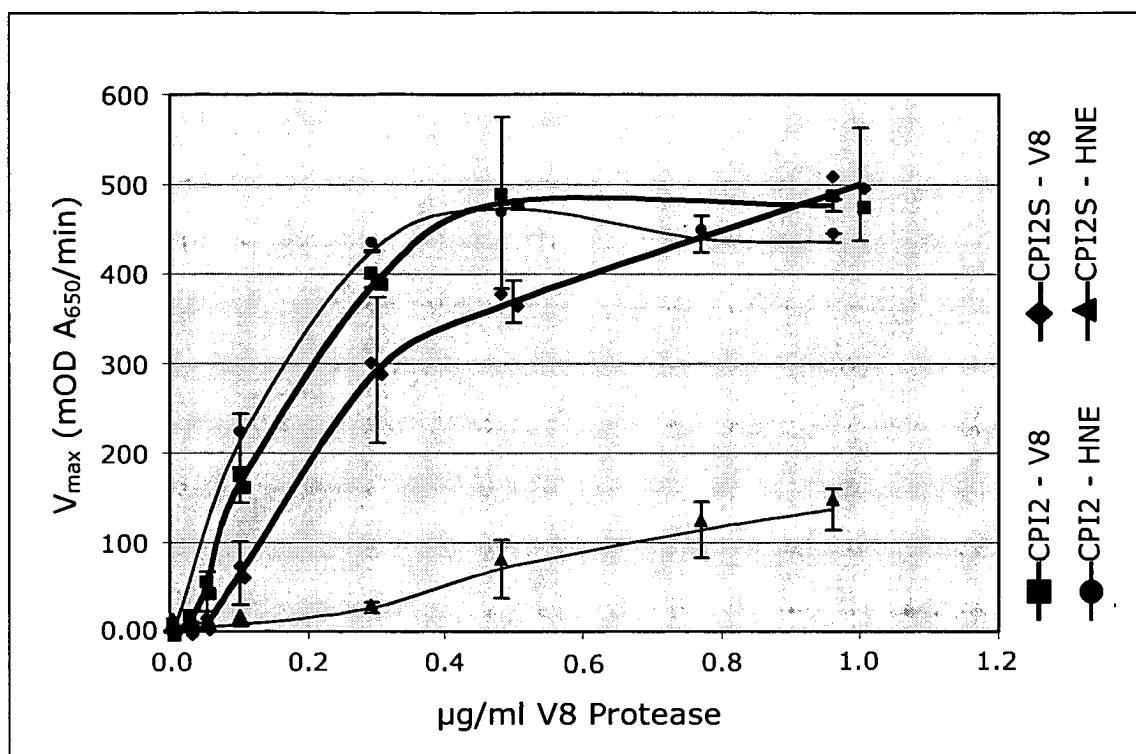


Figure 8

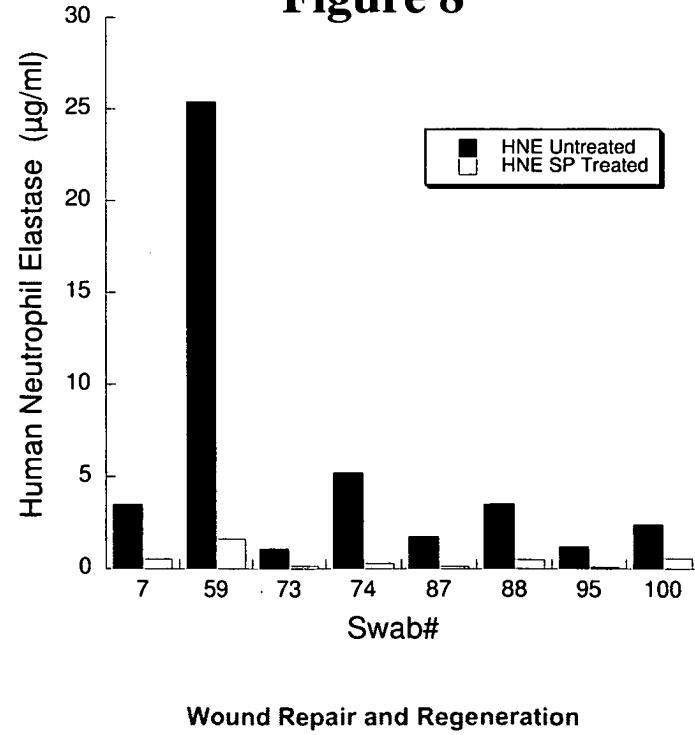


Figure 9

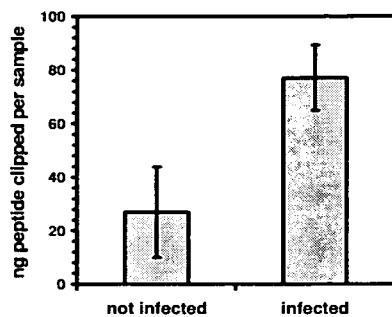


Figure 10 (a)

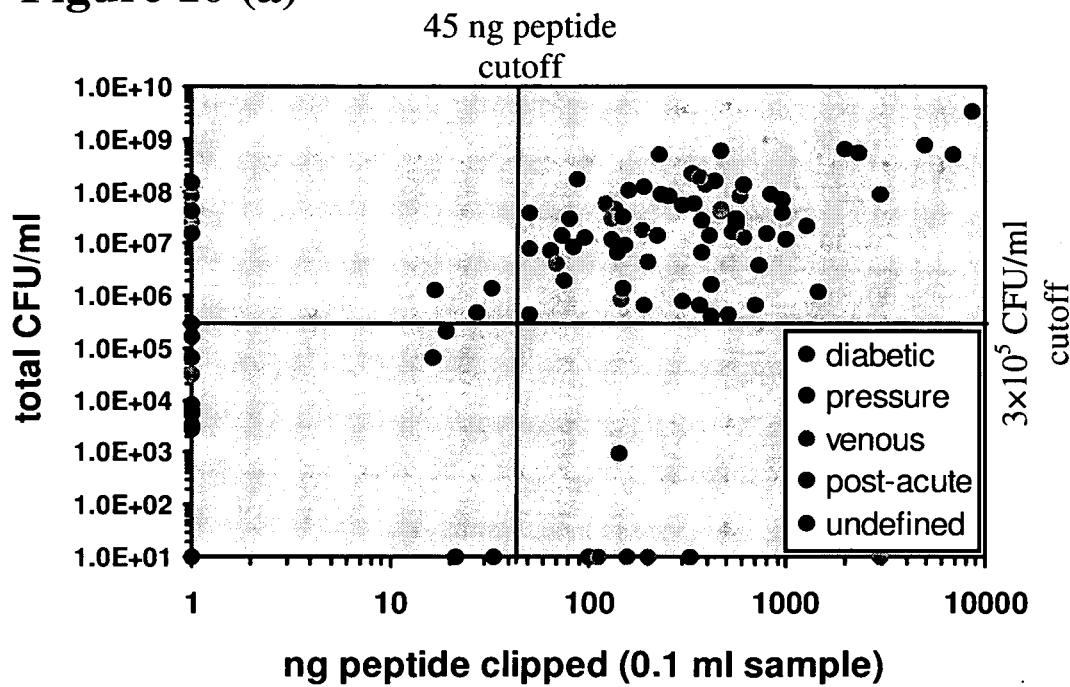


Figure 10 (b)

quantitative
microbiology

protease activity	+	-
	69	7

sensitivity = 89.6%
specificity = 69.6%

positive predictive value = 90.8%
negative predictive value = 66.7%

overall accuracy = 85.0%

Figure 11 (a)

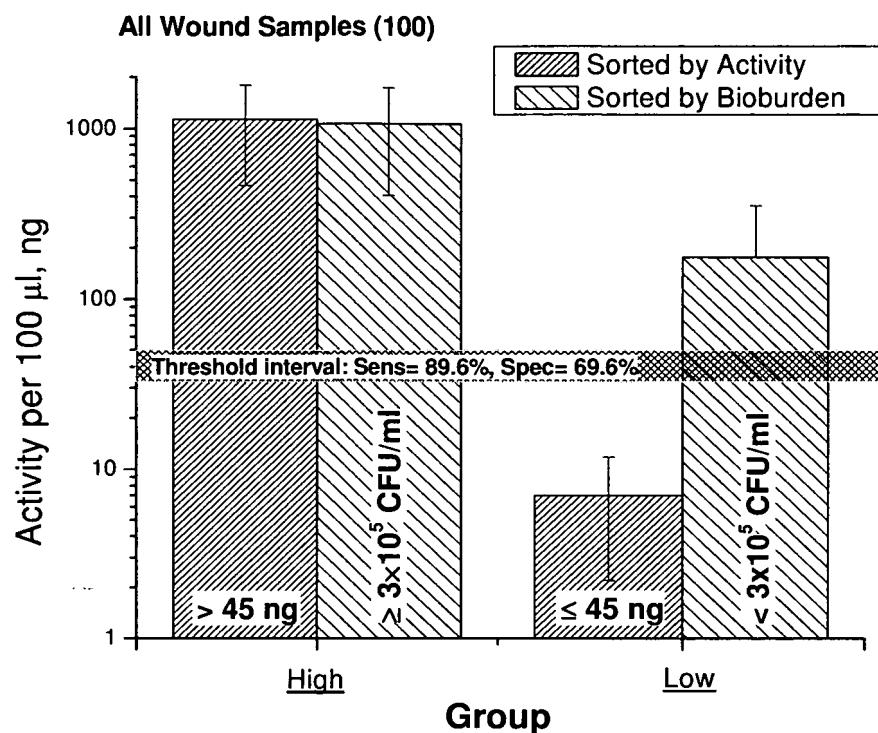


Figure 11 (b)

